



THE SPIDER ASSEMBLAGE IN A DENDROFLORISTIC HOTSPOT FROM EASTERN URUGUAY

Álvaro Laborda¹, Damián Hagopían^{1*}, Santiago Teijón¹, Joaquín Ginella¹, José Carlos Guerrero² & Miguel Simó¹

¹Sección Entomología. Facultad de Ciencias. Universidad de la República. Iguá 4225. CP 11400. Montevideo. Uruguay.

²Laboratorio de Desarrollo Sustentable y Gestión Ambiental del Territorio. Facultad de Ciencias. Universidad de la República. Iguá 4225. CP 11400. Montevideo. Uruguay.

*Corresponding author: dhagopian@fcien.edu.uy

ABSTRACT

In previous studies, Uruguay has been considered a biogeographical crossroads. Paso Centurión and Sierra de Ríos in eastern Uruguay were recently entered into the National System of Protected Areas. The landscape is characterized by a mosaic of different ecosystems located in one of the dendrofloristic hotspots proposed for the country. The spiders constitute a megadiverse group useful for the monitoring of natural environments. The aim of this study was to know the composition and structure of the spider fauna of this protected area. The sampling of spiders was carried out in two different environments (ravine and riparian forests) and three methods were used: ground vacuum, foliage beating and manual collection. A greater richness of spider families, species and guilds was recorded compared to previous studies in other protected areas from Uruguay. The two environments showed differences in taxonomic composition and species replacement. Araneidae, Theridiidae and Salticidae represented the families with the highest species richness. The family Symphytognathidae and 55 species are new records for the country. The finding of more southern records for several species confirm that the area is located in a border zone with other biogeographic units at regional level. This fact supports the crossroads condition for the country, provides key information for the management of the area and increases the number of priority spider species for conservation in Uruguay.

Keywords: Araneae, species list, guilds, protected area, conservation.

RESUMEN

El ensamble de arañas en un hotspot dendroflorístico del este de Uruguay. Estudios recientes han considerado a Uruguay como una encrucijada biogeográfica. Paso

Centurión y Sierra de Ríos en el este de Uruguay fue recientemente ingresado al Sistema Nacional de Áreas Protegidas. El paisaje está caracterizado por un mosaico de diferentes ecosistemas situado en uno de los *hotspots* dendroflorísticos propuestos para el país. Las arañas constituyen un grupo megadiverso que es útil para el control de los entornos naturales. El objetivo del estudio consistió en conocer la composición y estructura de la araneofauna de esta área protegida. El estudio se llevó a cabo en dos tipos de ambientes (bosque ribereño y bosque de quebrada) y se utilizaron tres métodos: aspirador de suelo, batido de follaje y recolección manual. Se registró una mayor riqueza de familias, especies y gremios de arañas en comparación con estudios previos en otras áreas protegidas de Uruguay. Los dos ambientes mostraron diferencias en la composición taxonómica y reemplazo de especies. Araneidae, Theridiidae y Salticidae fueron las familias con mayor riqueza específica. La familia Symphytognathidae y 55 especies son nuevos registros para el país. El hallazgo de registros más australes para varias especies confirma que esta área se encuentra en una zona límite con otras unidades biogeográficas a nivel regional. Esto refuerza el carácter de enclave biogeográfico para el país, aporta información relevante para el manejo del área y aumenta el número de especies prioritarias de arañas para la conservación en Uruguay.

Palabras clave: Araneae, lista de especies, gremios, área protegida, conservación.

INTRODUCTION

Biodiversity is essential for maintaining life and ecological services of ecosystems. In recent centuries, the process of habitat fragmentation, pollution, invasive species and the substitution of natural areas by different land uses have increased the rate of species extinction and the ecological interactions involved, causing a loss of biodiversity worldwide

(Ellis *et al.*, 2010; Valiente-Banuet *et al.*, 2015). These threats led to the need to establish natural protected areas for biota conservation (Le Saout *et al.*, 2013). One of the important tools for conservation is to describe the biodiversity and the study of ecological communities to understand their role in ecosystems (Norris, 2012). The hotspots are priority areas for conservation, at local and regional scales, because they are characterized by high species richness, the presence of endemic, rare or threatened species, high functional diversity, a unique evolutionary history and biogeography unit (Myers, 2003; Cañadas *et al.*, 2014; Marchese, 2015). The number of species has been the most common argument to apply in the conservation management of natural ecosystems, in particular plants to estimate endemism and vertebrates for comparison between areas. However, invertebrates have probably been poorly considered due to the scarcity of data (Marchese, 2015). Natural forests maintain a high floristic diversity and are key corridors for the gene flow of fauna and flora associated with these ecosystems (Dixo *et al.*, 2009; Dardengo *et al.*, 2016). Furthermore, the boundaries at which different biota converge are known as biogeographic crossroads. This confluence generates higher levels of alpha diversity and species replacement between environments or ecosystems (beta diversity), which is why they are considered priority areas of interest for conservation (Spector, 2002).

From a biogeographic point of view, Uruguay is included in the Pampean Province (Morrone, 2014) where grasslands are the most common landscape throughout the country. Grela (2004) established the first proposal of endemism and hotspots for the Uruguayan dendroflora and the connections with neighboring floristic biota in southern South America. He recognized four floristic hotspots in the country that reflect the influence with other biogeographic areas such as the Alto Paraná forest and the Chaco provinces. From this point of view, Uruguay constitutes a border zone between the temperate grasslands of the Pampas with different ecoregions (Olson *et al.*, 2001) or provinces (Morrone, 2014) of subtropical forests. Thus, the Uruguayan biogeographic regionalization proposed by Grela (2004) provided a framework for focusing future efforts on establishing protected areas in the country. Simó *et al.* (2014) reported the Uruguayan opiliofauna areas based on modeling of distribution species. They postulated this country as a biogeographic crossroads for the confluence of species from the Pampas and Paraná forests in agreement with Grela proposal for dendroflora. In addition, Laborda *et al.* (2018) showed the important role played by the riparian forests of the islands of the Uruguay River as a biological corridor for spider species with Alto Paraná forest. These facts place Uruguay as an area of high interest in the

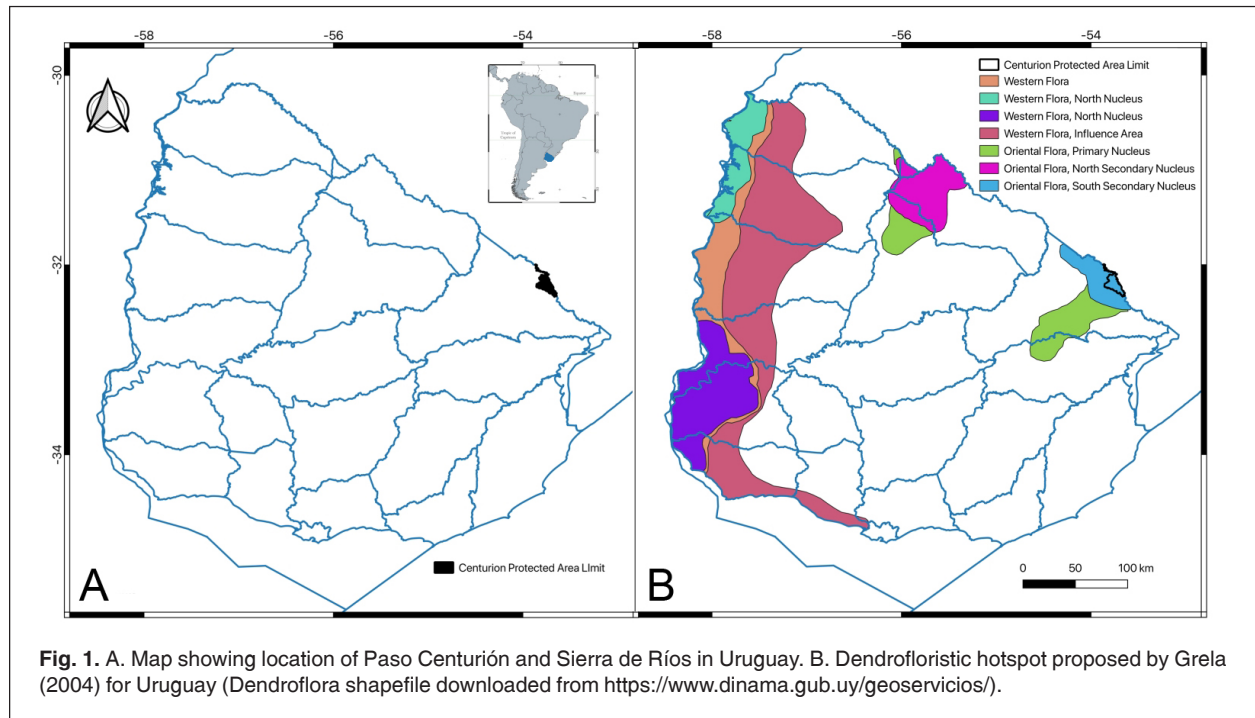
conservation of natural environments in South America. In spite of the uniqueness of the dendroflora, only 4.5% of the territory (Toranza *et al.* 2019) is represented by different types of natural forests such as riparian, hillside and ravine forests, which maintain biogeographic connections on a regional scale. Unfortunately, less than 1% of Uruguayan landscapes are included in the country's protected areas and, in addition, agricultural expansion and the loss of natural habitats for other productive activities have increased in recent decades (Brazeiro *et al.*, 2020). It is therefore imperative to generate knowledge about biodiversity as tools for planning new areas that cover more ecosystems and the corridors that connect them.

Spiders constitute a megadiverse group with 48770 species described worldwide (WSC 2020). Studies in South American riparian forests showed that these environments have a great richness of spider species, where spider diversity or the richness of spider guilds could be affected by vegetation structure (Rodrigues & Mendonça, 2012; Rodrigues *et al.*, 2014; Griotti *et al.*, 2017). Therefore, the high richness of spider species and guilds in Uruguay is expected to be associated with high plant diversity such as the four floristic hotspots proposed by Grela (2004). Efforts to record the spider diversity in Uruguay have been carried out over the last two decades, focusing on unexplored or little-known regions (Simó *et al.*, 2015, Laborda *et al.*, 2018). One of these regions is the east of the country, where Grela (2004) proposed two floristic hotspots. The northernmost where a rapid spider assessment was conducted in the truncated hills range (Simó *et al.* 2015). In this study, the authors found a high richness of spider species and proposed that the hills are linked to northern ecoregions based on the presence of species from Alto Parana forest and Araucaria forest. In the southernmost dendrofloristic hotspot, Paso Centurión and Sierra de Ríos were recently included in the National System of Protected Areas (SNAP, 2019). This area represents a mosaic of different ecosystems with high diversity and rare species (Grela, 2004; Brussa & Grela 2007; Brazeiro, 2015). Since knowledge about the spider fauna in this area is scarce, we conducted an assessment in order to study the spider community in one of the dendrofloristic hotspots from Uruguay as a baseline for the conservation management of the area.

MATERIAL AND METHODS

Study area

The study was carried out in Paso Centurión and Sierra de Ríos, in eastern Uruguay, near the border with Brazil (Fig. 1A). The area is located at the confluence of the Gondwananic Sedimentary Basin and Sierra del Este ecoregions (Brazeiro, 2015) and is connected to the Sul-Riograndense Planalto in



southern Brazil. It was considered one of the areas of eastern Uruguay with conservation priority based on that it was included in a dendrofloristic hotspot (Grela 2004) and because of the high species diversity in plants and vertebrates (Brazeiro, 2015) (Fig. 1B). The landscape is made up of three topographic units: 1) Highlands, which are characterized by hills with grasslands, shrublands and hillside forests, 2) Ravines, which are areas of steeply sloping terrain occupied by hillside forests, 3) Lowlands, with a predominance of plains with riparian forest along the rivers. The main productive activity in the area is extensive cattle raising and to a lesser extent crops, forestry, mineral production, wind energy and tourism (SNAP, 2019). For the study, we selected two sites: Paso Centurión ($32^{\circ}7'54.29''S$; $53^{\circ}44'5.24''W$) characterized by flood plains and riparian forests associated with the Yaguarón River and Sierra de Ríos ($32^{\circ}11'0.79''S$; $53^{\circ}51'15.02''W$) composed of hills with ravine forests and grasslands associated with rock formations.

Sampling procedure

Seasonal samplings were conducted in July (2016, winter), April (2017 and 2019, autumn) and November (2017, spring) in two different environments: riparian forests and ravine forests, each associated with grassland and rocky areas ecotones (Fig. 2). Three complementary collecting methods were used in each survey: ground vacuum

for ground and herbaceous vegetation stratum, consisting of 20 daytime and 20 one-minute nighttime samples; manual collection, four one-hour daytime and four one-hour nighttime samples; and foliage beating consisting of 20 daytime samples, each represented by 15 hits on the tree vegetation collected in 1 m^2 of a white cloth. These methods are considered very effective in capturing spiders from foliage and soil (Coddington *et al.* 1996; Pérez-Miles *et al.* 1999; Adis 2002; Schmidt *et al.* 2006; Jorge *et al.*, 2013). Juveniles and adults were identified at the family level in order to recognize functional guilds. Adults were identified for morphospecies as far as possible at the species level through the use of keys (Jocqué & Dippenaar-Schoeman, 2007; Grismado *et al.*, 2014) and taxonomic reviews. Spider guilds were classified according to Cardoso *et al.* (2011). We collected under DINAMA scientific collecting permit No. 2020/14000/003764. The voucher specimens were deposited in the arachnological collection of Facultad de Ciencias, Universidad de la República, Uruguay. A map of the study area was elaborated using QGIS (2020).

Statistics

To test the statistical differences in taxonomic composition between the two surveyed sites, we conducted an ANOSIM test. In order to evaluate the contribution of the species to the similarity/dissimilarity

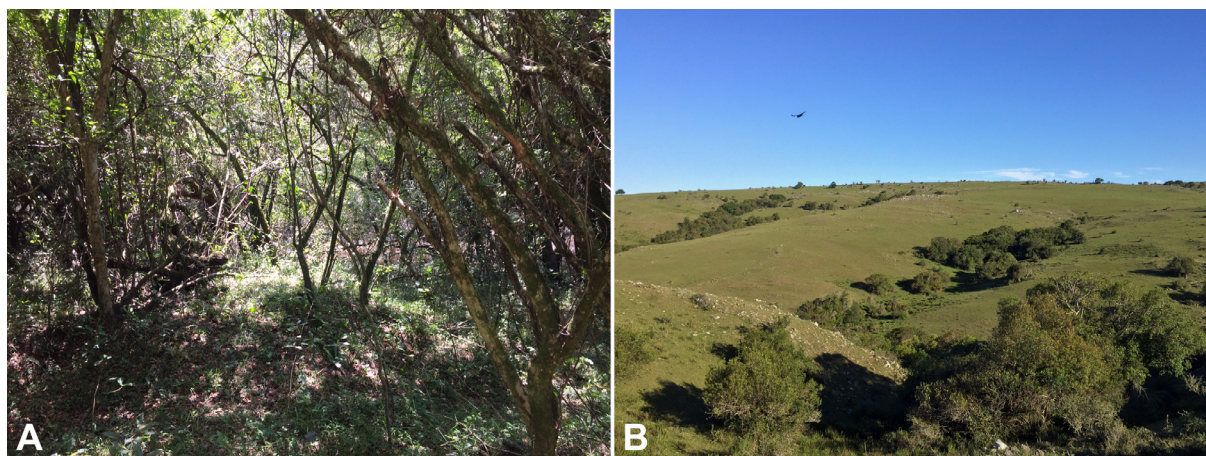


Fig. 2. Paso Centuri3n and Sierra de R3os. Environments studied. A. Riparian forest, B. Ravine forest.

between the environments, we performed a SIMPER test. The statistics were calculated using PAST 4.03 software (Hammer *et al.*, 2001).

RESULTS

We collected 2591 spiders, represented by 38 families, 127 genera and 252 species/morphospecies (Table 1). One family (Symphytognathidae), 30 genera and 55 species are new records for Uruguay (Figs. 3 and 4). Furthermore, 60% were juveniles, 25% adult females and 15% adult males. Almost all of the species recorded (72%) belong to seven families, Araneidae (18%), Theridiidae (17%), Salticidae (14%), Linyphiidae (8%), Lycosidae (6%), Thomisidae (5%) and Anyphaenidae (4%). The number of species recorded was higher in the riparian forest ($S = 206$) than in the ravine forest ($S = 83$). More than half of the species recorded belong to the guilds of Other Hunters (24%), Orb Web (23%), and Space Web (19%).

Species composition for the environments studied (ravines and riparian forest) were significantly different (ANOSIM test: Bray-Curtis index $R=0.1054$, $p=0.0001$; Morisita index $R=0.1074$, $p=0.0001$) (Fig. 5) and with a high degree of species replacement (Jaccard similarity index: 0.146825). According to the SIMPER test of comparison between environments, *Faiditus plaumanni* (Theridiidae) was the species that most contributed to the observed dissimilarity (5.3%). This species was found in both types of environments studied, but with greater abundance in the riparian forest. All other species have less than 5% of contribution.

DISCUSSION

The high number of new spider records for the country increased the knowledge of the Uruguayan spider fauna. In addition, the high species richness and endemism of this area confirmed Paso Centuri3n and Sierra de R3os as a hotspot for the spiders in eastern Uruguay. The results indicate that this area has the highest spider species richness compared to other natural environments studied in the country (Sim3 *et al.*, 1994; Costa *et al.*, 2006; Sim3 *et al.*, 2015; Laborda *et al.*, 2018). The dendroflora is influenced by the southern limit of the Brazilian Atlantic Forest (Brussa & Grela, 2007) that constitutes the Paso Centuri3n and Sierra de R3os as the biotic limit of this ecoregion with the Pampas, which provides additional interest for conservation. This is supported by several species of spiders found here ($S = 46$; 18%) where their presence represents the southernmost record. In addition, these species agree with six of the seven criteria proposed by Ghione *et al.* (2017) for considering arachnid species as a conservation priority in Uruguay. They are as follows: C1, species endemic to Uruguay; C2, species with few records in arachnological collections in the country; C3, species only present in threatened environments; C4, species only recorded in rare habitats in the country; C5, species in which Uruguay represents the southernmost record in its distribution area; C6, species with ecological uniqueness. This last criterion was assumed on the basis that these species are associated with subtropical forests that encroach on Uruguayan territory (Brussa & Grela, 2007). As a result of this study, 12 species of those indicated by

Table 1. Taxonomic list of spiders collected in Paso Centurión and Sierra de Ríos (protected area), indicating guilds and associated environments for each species. New records are indicated with an asterisk (*). Southernmost records are indicated with a superscript S (^S). New species to be considered with conservation priority for Uruguay are indicated with a superscript NPC (^{NPC}). Species considered with conservation priority for Uruguay *sensu* Ghione *et al.* (2015) are indicated with a superscript PC (^{PC}). Abbreviations: **AH** = Ambush Hunters; **GH** = Ground Hunters; **OH** = Other Hunters; **OW** = Orb Web; **R** = Ravine; **RF** = Riparian Forest; **S** = Specialists; **ShH** = Sheet Hunters; **ShW** = Sheet Web; **SpW** = Space Web; **SW** = Sensing Web.

	Guild	Environment		Guild	Environment
Actinopodidae			<i>Micrathena spitzii</i> Mello-Leitão, 1932* ^{SNPC}	OW	RF/R
<i>Actinopus</i> sp.	SW	R	<i>Mecynogea bigibba</i> Simon, 1903	OW	RF
Amaurobiidae			<i>Metepeira compsa</i> (Chamberlin, 1916)* ^{SNPC}	OW	RF
Amaurobiidae sp.1	ShW	RF	<i>Ocrepeira gnomo</i> (Mello-Leitão, 1943)* ^{SNPC}	OW	RF
Amaurobiidae sp.2	ShW	R	<i>Ocrepeira</i> sp.	OW	RF
<i>Retiro nigrionotatus</i> Mello-Leitão, 1947* ^{SNPC}	ShW	RF/R	<i>Ocrepeira venustula</i> (Keyserling, 1879)	OW	RF/R
Anyphaenidae			<i>Paraverrucosa heteracantha</i> (Mello-Leitão, 1943)*	OW	RF
Anyphaenidae sp.1	OH	RF	<i>Parawixia audax</i> (Blackwall, 1863)	OW	RF/R
<i>Aysha</i> sp.1	OH	R	<i>Parawixia undulata</i> (Keyserling, 1892)	OW	RF
<i>Aysha</i> sp.2	OH	R	<i>Trichonephila clavipes</i> (Linnaeus, 1767)	OW	RF
<i>Aysha</i> sp.3	OH	RF	<i>Verrucosa meridionalis</i> (Keyserling, 1892)* ^{SNPC}	OW	RF/R
<i>Aysha</i> sp.4	OH	RF	<i>Verrucosa undecimvariolata</i>		
<i>Aysha borgmeyeri</i> (Mello-Leitão, 1926)* ^{SNPC}	OH	RF	(O. Pickard-Cambridge, 1889)	OW	RF
<i>Aysha vacaria</i> Brescovit, 1992* ^{SNPC}	OH	RF	<i>Wagneriana atuna</i> Levi, 1991* ^{SNPC}	OW	RF
RF/R			Caponiidae		
<i>Sanogasta backhauseni</i> (Simon, 1895)	OH	R	<i>Nops meridionalis</i> Keyserling, 1891* ^{SNPC}	S	R
<i>Tasata variolosa</i> Mello-Leitão, 1943	OH	RF	Cheiracanthiidae		
<i>Xiruana</i> sp.	OH	RF	<i>Cheiracanthium inclusum</i> (Hentz, 1847)	OH	RF
Araneidae			<i>Eutichurus ibiuna</i> Bonaldo, 1994	OH	RF
<i>Acacesia tenella</i> (L. Koch, 1871)* ^{SNPC}	OW	RF	Clubionidae		
<i>Alpaida carminea</i> (Taczanowski, 1878)	OW	R	<i>Elaver brevipes</i> (Keyserling, 1891)* ^{SNPC}	OH	R
<i>Alpaida ericae</i> Levi, 1988*	OW	RF	Corinnidae		
<i>Alpaida rubellula</i> (Keyserling, 1892)	OW	RF	<i>Castianeira</i> sp.1	GH	R
<i>Alpaida veniliae</i> (Keyserling, 1865)	OW	R	<i>Castianeira</i> sp.2	GH	RF
<i>Alpaida</i> sp.	OW	RF	<i>Castianeira</i> sp.3	GH	R
<i>Araneus lathyrinus</i> (Holmberg, 1875)	OW	R	<i>Castianeira</i> sp.4	GH	RF
<i>Araneus omnicolor</i> (Keyserling, 1893)	OW		<i>Falconina gracilis</i> (Keyserling, 1891)	GH	R
RF/R			Ctenidae		
<i>Araneus unifromis</i> (Keyserling, 1879)	OW	RF/R	<i>Asthenoctenus borellii</i> Simon, 1897	OH	RF
<i>Araneus workmani</i> (Keyserling, 1884)	OW	RF	<i>Guasuctenus longipes</i> (Keyserling, 1891) ^{PC}	OH	RF/R
<i>Argiope argentata</i> (Fabricius, 1775)	OW	RF	<i>Parabatinga brevipes</i> (Keyserling, 1891)	OH	RF
<i>Cyclosa fililineata</i> Hingston, 1932* ^{SNPC}	OW	RF	Desidae		
<i>Cyclosa machadinho</i> Levi, 1999	OW	RF/R	<i>Meteltella iheringhi</i> (Keyserling, 1891)	ShW	RF
<i>Cyclosa morretes</i> Levi, 1999* ^{SNPC}	OW	RF/R	<i>Metaltella simoni</i> (Keyserling, 1878)	ShW	RF
<i>Eriophora</i> sp. *	OW	RF	Filistatidae		
<i>Eustala cidae</i> Poeta, 2014* ^{SNPC}	OW	RF	<i>Pikelinia arenicola</i> Lise,		
<i>Eustala saga</i> (Keyserling, 1893)	OW	RF/R	Ferreira & Silva, 2010* ^{SNPC}	ShW	RF
<i>Eustala minuscula</i> (Keyserling, 1892)	OW	RF	Gnaphosidae		
<i>Eustala</i> sp.1	OW	RF	<i>Almafuerie</i> sp.	GH	RF
<i>Eustala</i> sp.2	OW	RF	<i>Apodrassodes guatemalensis</i>		
<i>Eustala</i> sp.3	OW	RF/R	(F. O. Pickard-Cambridge, 1899)	GH	RF
<i>Eustala taquara</i> (Keyserling, 1892)	OW	R	<i>Apopyllus atlanticus</i> Azevedo, Ott,		
<i>Gasteracantha cancriformis</i> (Linnaeus, 1758)	OW	RF/R	Griswold & Santos, 2016* ^{SNPC}	GH	RF
<i>Kaira altiventer</i> O. Pickard-Cambridge, 1889* ^{SNPC}	OW	RF	<i>Camillina galianoae</i> Platnick & Murphy, 1987	GH	RF
<i>Larinia bivittata</i> Keyserling, 1885	OW	RF	Gnaphosidae sp.	GH	RF
<i>Mangora fundo</i> Levi, 2007	OW	RF/R	<i>Latonigena auricomis</i> Simon, 1893	GH	RF
<i>Mangora strenua</i> (Keyserling, 1893)* ^{SNPC}	OW	RF/R	Hahniidae		
<i>Mastophora catarina</i> Levi, 2003* ^{SNPC}	OW	RF	Hahniidae sp.	ShW	RF
<i>Mastophora</i> sp.	OW	R	Linyphiidae		
<i>Micrathena furva</i> (Keyserling, 1892)	OW	RF/R	<i>Dubiaranea difficilis</i> (Mello-Leitão, 1944)	ShH	R
<i>Micrathena furcata</i> (Hahn, 1822)	OW	RF	Linyphiidae sp.1	ShH	R
<i>Micrathena nigrichelis</i> Strand, 1908	OW	R	Linyphiidae sp.2	ShH	R

Table 1. Cont.

	Guild	Environment		Guild	Environment
Linyphiidae sp.3	ShH	RF	<i>Asaphobelis physonychus</i> Simon, 1902* ^{SNPC}	OH	RF
Linyphiidae sp.4	ShH	R	<i>Breda bicrucata</i> (Mello-Leitão, 1943)	OH	RF
Linyphiidae sp.5	ShH	RF	<i>Breda tristis</i> Mello-Leitão, 1944	OH	R
Linyphiidae sp.6	ShH	RF	<i>Chira aff. thysbe</i> Simon, 1902*	OH	RF
Linyphiidae sp.7	ShH	RF	<i>Colonus melanogaster</i> (Mello-Leitão, 1917)* ^{SNPC}	OH	RF
Linyphiidae sp.8	ShH	RF	<i>Cotinusa melanura</i> Mello-Leitão, 1939* ^{SNPC}	OH	RF
Linyphiidae sp.9	ShH	RF	<i>Cotinusa trifasciata</i> (Mello-Leitão, 1943)	OH	RF
Linyphiidae sp.10	ShH	RF	<i>Dendryphantini</i> sp.1	OH	RF
Linyphiidae sp.11	ShH	RF	<i>Dendryphantini</i> sp.2	OH	RF
Linyphiidae sp.12	ShH	R	<i>Dendryphantini</i> sp.3	OH	RF
Linyphiidae sp.13	ShH	RF	<i>Dendryphantini</i> sp.4	OH	RF
Linyphiidae sp.14	ShH	RF/R	<i>Euophryini</i> sp.1	OH	RF
Linyphiidae sp.15	ShH	RF	<i>Euophryini</i> sp.2	OH	RF
Linyphiidae sp.16	ShH	RF	<i>Euophrys melanoleuca</i> Mello-Leitão, 1944	OH	RF
<i>Scolecurea cambara</i> Rodrigues, 2005* ^{SNPC}	GH	RF/R	<i>Gastromicans cf. vigena</i>		
<i>Scolecurea</i> sp.	GH	RF	(Peckham & Peckham, 1901)*	OH	RF
<i>Sphecozone ignigena</i> (Keyserling, 1886)	GH	R	<i>Lyssomanes pauper</i> Mello-Leitão, 1945	OH	
<i>Sphecozone</i> sp.	GH	R	RF/R		
Lycosidae			<i>Megafreya sutrix</i> (Holmberg, 1875)	OH	RF
<i>Agalenocosa</i> sp.	GH	R	<i>Metaphidippus</i> sp.*	OH	RF
<i>Agalenocosa velox</i> (Keyserling, 1891)	GH	RF	<i>Neonella lubrica</i> Galiano, 1988* ^{SNPC}	OH	RF
<i>Aglaoctenus lagotis</i> (Holmberg, 1876) ^{PC}	GH	RF	<i>Neonella montana</i> Galiano, 1988*	OH	RF
<i>Aglaoctenus oblongus</i> (C. L. Koch, 1847) ^{PC}	GH	RF	<i>Neonella</i> sp.	OH	RF
<i>Diapontia uruguayensis</i> Keyserling, 1877	GH	RF	<i>Philira micans</i> (Simon, 1902)* ^{SNPC}	OH	RF
<i>Hogna gumia</i> (Petrunkevitch, 1911)* ^{SNPC}	GH	RF/R	<i>Pseudofluda pulcherrima</i> Mello-Leitão, 1928* ^{SNPC}	OH	RF
<i>Lobizon humilis</i> (Mello-Leitão, 1944)	GH	RF/R	<i>Saphrys saitiformis</i> (Simon, 1901)	OH	RF
<i>Lycosa erythrognatha</i> Lucas, 1836	GH	RF	<i>Sarinda marcosi</i> Piza, 1937	OH	RF
<i>Lycosa poliostoma</i> (C. L. Koch, 1847)	GH	RF/R	<i>Sumampattus hudsoni</i> Galiano, 1996	OH	RF
Lycosinae sp.1	GH	RF/R	<i>Tartamura adfectuosa</i> (Galiano, 1977)*	OH	RF
Lycosinae sp.2	GH	R	<i>Therrella galianoae</i> Brul & Lise, 1996* ^{SNPC}	OH	RF
Lycosinae sp.3	GH	RF	<i>Titanattus andinus</i> (Simon, 1900)	OH	R
<i>Navira nagan</i> Piacentini & Grismado, 2009	GH	R	<i>Tullgrenella morenensis</i> (Tullgren, 1905)* ^{SNPC}	OH	RF
<i>Paratrochosina amica</i> (Mello-Leitão, 1941)*	GH	RF	<i>Tullgrenella musica</i> (Mello-Leitão, 1945)*	OH	RF
<i>Pavocosa gallopavo</i> (Mello-Leitão, 1941)	GH	RF	<i>Tullgrenella serrana</i> Galiano, 1970	OH	R
Mimetidae			<i>Tullgrenella</i> sp.	OH	R
<i>Gelanor zonatus</i> (C. L. Koch, 1845) ^{PC}	S	RF	<i>Zygoballus</i> sp.*	OH	RF
<i>Mimetus melanoleucus</i> Mello-Leitão, 1929	S	R	Scytodidae		
Miturgidae			<i>Scytodes globula</i> Nicolet, 1849	OH	RF/R
<i>Teminius insularis</i> (Lucas, 1857)	OH	RF	Segestriidae		
Mysmenidae			<i>Ariadna boesenbergi</i> Keyserling, 1877	SW	RF
<i>Microdipoena</i> sp.	SpW	R	Selenopidae		
Mysmenidae sp.	SpW	RF	<i>Selenops rapax</i> Mello-Leitão, 1929* ^{SNPC}	AH	RF
Oonopidae			<i>Selenops spixi</i> Perty, 1833	AH	RF
<i>Neotrops lorenae</i> Grismado & Ramírez, 2013	GH	RF	Senoculidae		
Oonopidae sp.	GH	RF	<i>Senoculus purpureus</i> (Simon, 1880)* ^{SNPC}	OH	RF
<i>Orchestina</i> sp.	GH	RF	Sparassidae		
Oxyopidae			<i>Polybetes pallidus</i> Mello-Leitão, 1941	OH	RF
<i>Hamataliwa cf. marmorata</i> Simon, 1898	OH	RF	<i>Polybetes punctulatus</i> Mello-Leitão, 1944	OH	RF
<i>Oxyopes salticus</i> Hentz, 1845	OH	RF	<i>Polybetes pythagoricus</i> (Holmberg, 1875)	OH	RF
Palpimanidae			<i>Polybetes rapidus</i> (Keyserling, 1880)	OH	R
<i>Otiotrops birabeni</i> Mello-Leitão, 1945 ^{PC}	S	R	Symphytognathidae*		
Pholcidae			<i>Anapistula</i> sp.*	OW	R
<i>Mesabolivar charrua</i> Machado, Laborda, Simó & Brescovit, 2013 ^{PC}	SpW	RF/R	Tetragnathidae		
Pycnothelidae			<i>Chrysmeta boraceia</i> Levi, 1986	OW	RF/R
<i>Acanthogonathus tacuariensis</i>			<i>Leucauge volupis</i> (Keyserling, 1893)	OW	RF/R
(Pérez-Miles & Capocasale, 1982) ^{PC}	SW	R	<i>Leucauge</i> sp.	OW	R
Salticidae			<i>Tetragnatha</i> sp.1	OW	RF
<i>Aphirape flexa</i> Galiano, 1981	OH	RF/R	<i>Tetragnatha</i> sp.2	OW	RF

Table 1. Cont.

	Guild	Environment		Guild	Environment
Theraphosidae					
<i>Catumiri parvum</i> (Keyserling, 1878)	SW	RF			
Theridiidae					
<i>Achaearanea</i> sp. *	SpW	RF	Theridiidae sp.3	SpW	RF
<i>Anelosimus ethicus</i> (Keyserling, 1884)* ^{SNPC}	SpW	RF	Theridiidae sp.4	SpW	RF
<i>Anelosimus</i> sp.	SpW	RF	Theridiidae sp.5	SpW	RF
<i>Anelosimus viera</i> Agnarsson, 2012 ^{PC}	SpW	RF	<i>Theridion calcynatum</i> Holmberg, 1876	SpW	RF
<i>Argyrodes elevatus</i> Taczanowski, 1873	SpW	R	<i>Theridion</i> sp.1	SpW	RF
<i>Argyrodes</i> sp.	SpW	RF	<i>Theridion</i> sp.2	SpW	RF
<i>Ariamnes longissimus</i> Keyserling, 1891* ^{SNPC}	SpW	RF	<i>Thymoites puer</i> (Mello-Leitão, 1941)	SpW	RF
<i>Coleosoma acutiventer</i> (Keyserling, 1884)*	SpW	RF	<i>Wamba crispulus</i> (Simon, 1895)*	SpW	RF
<i>Cryptachaea altiventer</i> (Keyserling, 1884)	SpW	RF/R	Theridiosomatidae		
<i>Cryptachaea belulla</i> (Keyserling, 1891)	SpW	R	Theridiosomatidae sp.1	OW	RF
<i>Cryptachaea ericae</i> Rodrigues & Poeta, 2015* ^{SNPC}	SpW	RF	Theridiosomatidae sp.2	OW	RF
<i>Cryptachaea</i> sp.1	SpW	RF	Thomisidae		
<i>Cryptachaea</i> sp.2	SpW	RF	<i>Epicadinus villosus</i> Mello-Leitão, 1929 ^{SNPC}	AH	RF
<i>Cryptachaea</i> sp.3	SpW	RF	<i>Epicadus rubripes</i> Mello-Leitão, 1924* ^{NPC}	AH	RF
<i>Cryptachaea</i> sp.4	SpW	RF	<i>Misumenops pallidus</i> (Keyserling, 1880)	AH	RF
<i>Cryptachaea</i> sp.5	SpW	RF	<i>Strophius albofasciatus</i> Mello-Leitão, 1929	AH	RF
<i>Dipoena granulata</i> (Keyserling, 1886)	SpW	RF/R	<i>Titidius</i> sp.1	AH	RF
<i>Dipoena santacatarinae</i> Levi, 1963* ^{SNPC}	SpW	RF	<i>Titidius</i> sp.2	AH	RF
<i>Dipoena</i> sp.1	SpW	RF	<i>Titidius</i> sp.3	AH	RF
<i>Dipoena</i> sp.2	SpW	RF	<i>Titidius</i> sp.4	AH	R
<i>Dipoena</i> sp.3	SpW	R	<i>Titidius</i> sp.5	AH	RF
<i>Dipoena variabilis</i> (Keyserling, 1886)* ^{SNPC}	SpW	RF	<i>Tmarus cf. albifrons</i> Piza, 1944	AH	RF/R
<i>Faiditus americanus</i> (Taczanowski, 1874)*	SpW	RF/R	<i>Tmarus</i> sp.1	AH	RF
<i>Faiditus altus</i> (Keyserling, 1891)* ^{SNPC}	SpW	RF	<i>Tmarus</i> sp.2	AH	R
<i>Faiditus plaumanni</i> (Exline & Levi, 1962)* ^{SNPC}	SpW	RF/R	Trachelidae		
<i>Faiditus sicki</i> (Exline & Levi, 1962)* ^{SNPC}	SpW	RF	<i>Meriola</i> sp.1	GH	RF
<i>Faiditus</i> sp.1	SpW	RF	<i>Meriola</i> sp.2	GH	RF
<i>Faiditus</i> sp.2	SpW	RF	Trachelidae sp.1	GH	RF
<i>Guaraniella manherti</i> Baert, 1984	SpW	RF	Trachelidae sp.2	GH	R
<i>Rhomphaea brasiliensis</i> Mello-Leitão, 1920* ^{SNPC}	SpW	RF	Trachelidae sp.3	GH	R
<i>Rhomphaea paradoxa</i> (Taczanowski, 1873)* ^{SNPC}	SpW	RF	<i>Trachelopachys keyserlingi</i> (Roewer, 1951)* ^{SNPC}	GH	R
<i>Styopsis selis</i> Levi, 1964* ^{SNPC}	SpW	RF	Trechaleidae		
<i>Steatoda</i> sp.	SpW	RF	<i>Paratrechalea ornata</i> (Mello-Leitão, 1943) ^{PC}	S	RF/R
<i>Tekellina</i> sp. *	SpW	RF	<i>Trechaleoides biocellata</i> (Mello-Leitão, 1926) ^{PC}	S	R
Theridiidae sp.1	SpW	RF	Uloboridae		
Theridiidae sp.2	SpW	RF	<i>Conifaber yasi</i> Grismado, 2004* ^{SNPC}	OW	RF
			<i>Miagrammopes guttatus</i> Mello-Leitão, 1937* ^{SNPC}	OW	RF/R
			<i>Miagrammopes</i> sp.	OW	RF
			<i>Uloborus elongatus</i> Opell, 1982	OW	RF/R
			<i>Uloborus</i> sp.	OW	RF

Ghione *et al.* (2015) were found in the protected area and 46 new species were categorized here, thus increasing the number with conservation priority for Uruguay from 35 to 81 spider species.

The family Symphytognathidae was first registered in Uruguay. This family comprises very small spiders, mainly extended in the tropics, which build orb webs in the litter (Jocqué & Dippenaar-Schoeman, 2007). The genus *Anapistula* Gertsch, 1941 is represented in the Neotropical Region with nine species described. Some of them have been recorded for the Atlantic Forest in southern Brazil

and the Yungas in Argentina (Rheims & Brescovit, 2003; Rubio & González, 2010). The species collected in Paso Centurión and Sierra de Ríos are not known; so they must be described.

Araneidae, Theridiidae and Salticidae showed high species richness at the family level. Araneidae include orb-web weavers, Theridiidae build three-dimensional webs, and Salticidae include jumping spiders that are foliage or ground stalking hunters (Cardoso *et al.*, 2011). Representatives of these families are reported to be very abundant and diverse in the Atlantic Forest (Rodrigues *et al.*, 2014; Rubio,

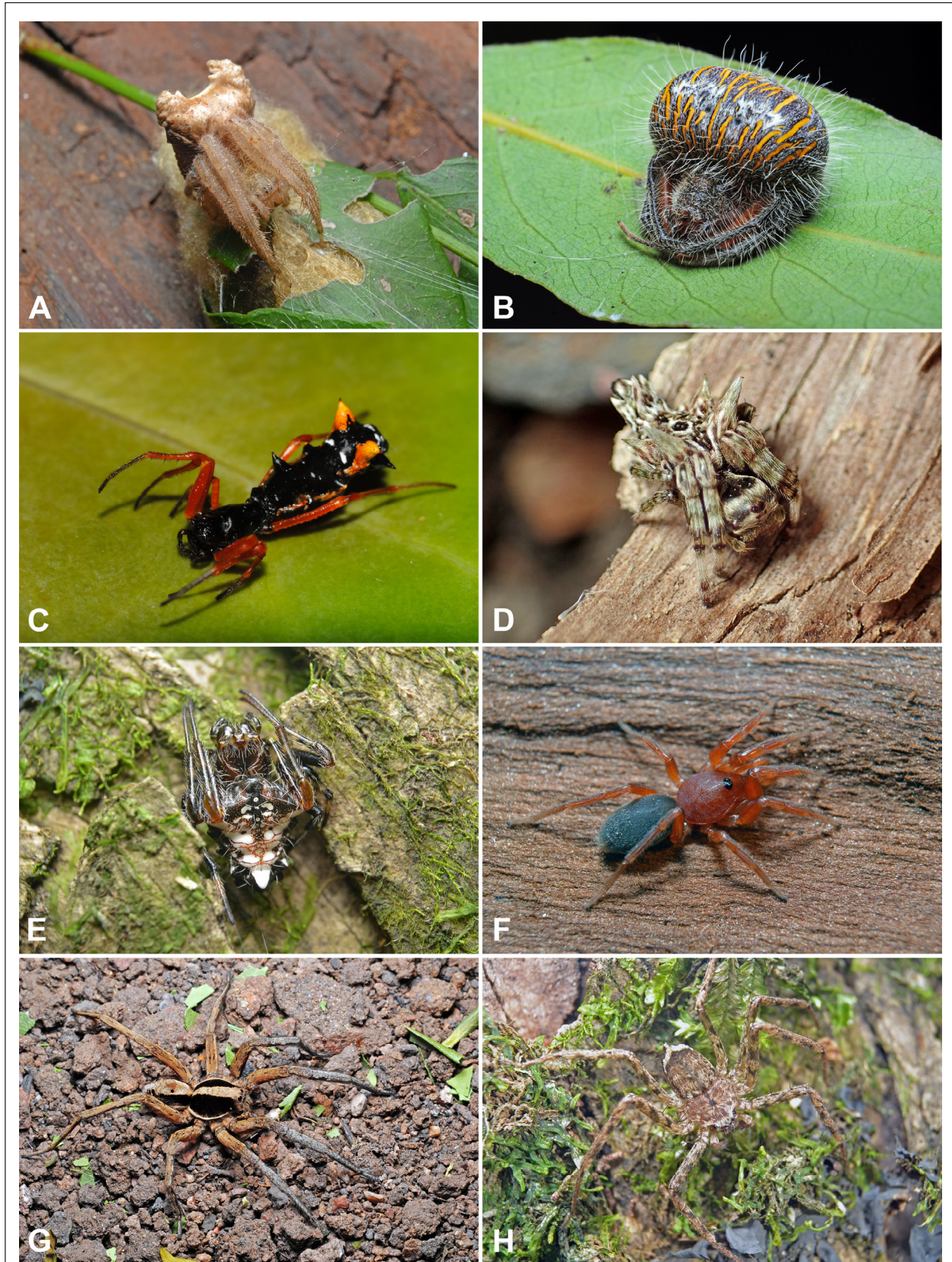


Fig. 3. New spider records of Paso Centuri3n and Sierra de R3os I. A. *Kaira altiventer* (Araneidae). B. *Mastophora catarina* (Araneidae). C. *Micrathena spitzii* (Araneidae). D. *Paraverrucosa heteracantha* (Araneidae). E. *Verrucosa meridionalis* (Araneidae). F. *Nops meridionalis* (Caponiidae). G. *Hogna gumia* (Lycosidae). H. *Selenops rapax* (Selenopidae).

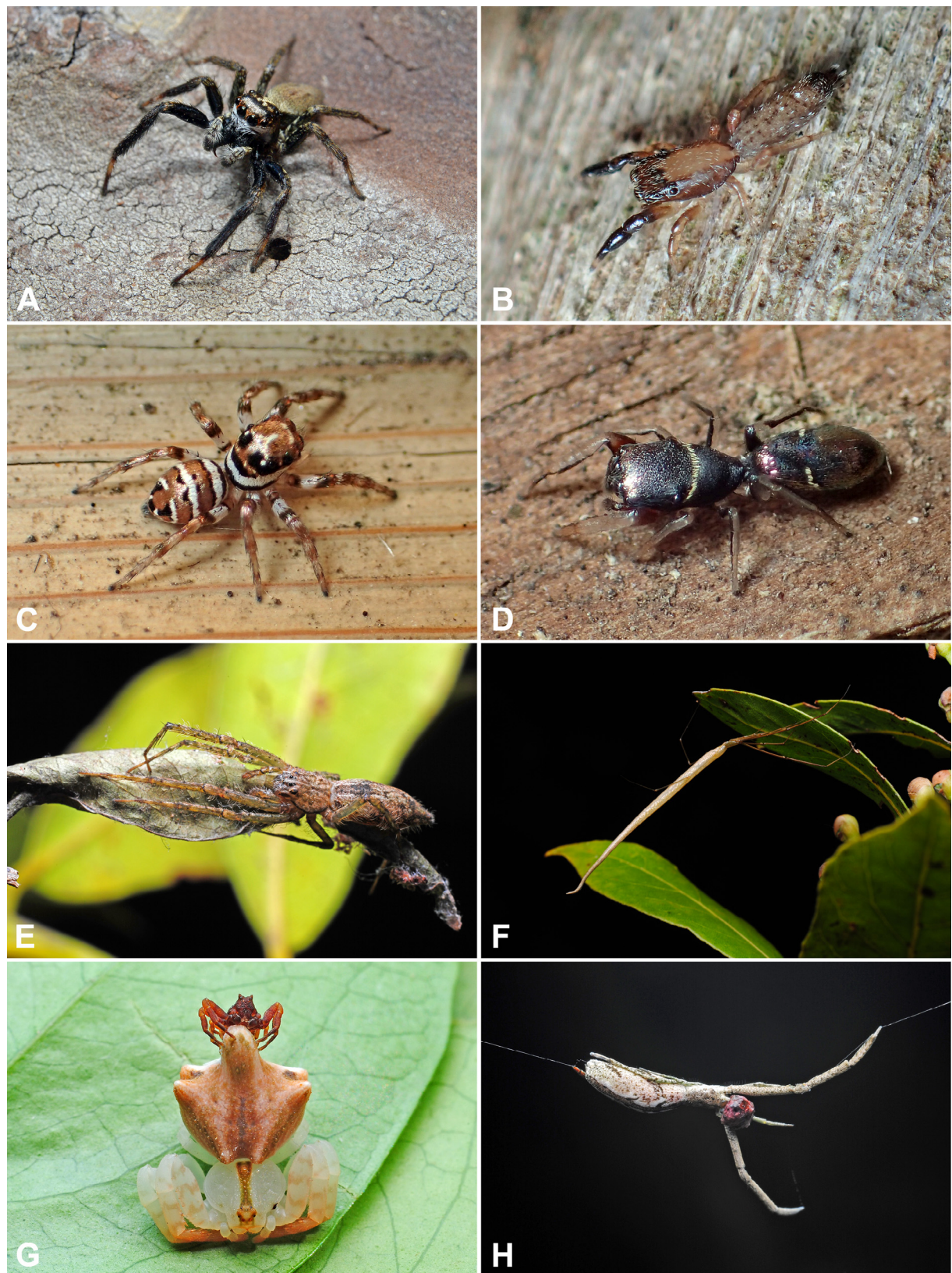


Fig. 4. New spider records of Paso Centurión and Sierra de Ríos II. A. *Asaphobelis physonychus* (Salticidae). B. *Cotinusa melanura* (Salticidae). C. *Philira micans* (Salticidae). D. *Pseudofluda pulcherrima* (Salticidae). E. *Senoculus purpureus* (Senoculidae). F. *Ariamnes longissimus* (Theridiidae). G. *Epicadus rubripes* (Thomisidae). H. *Miagrammopes guttatus* (Uloboridae).

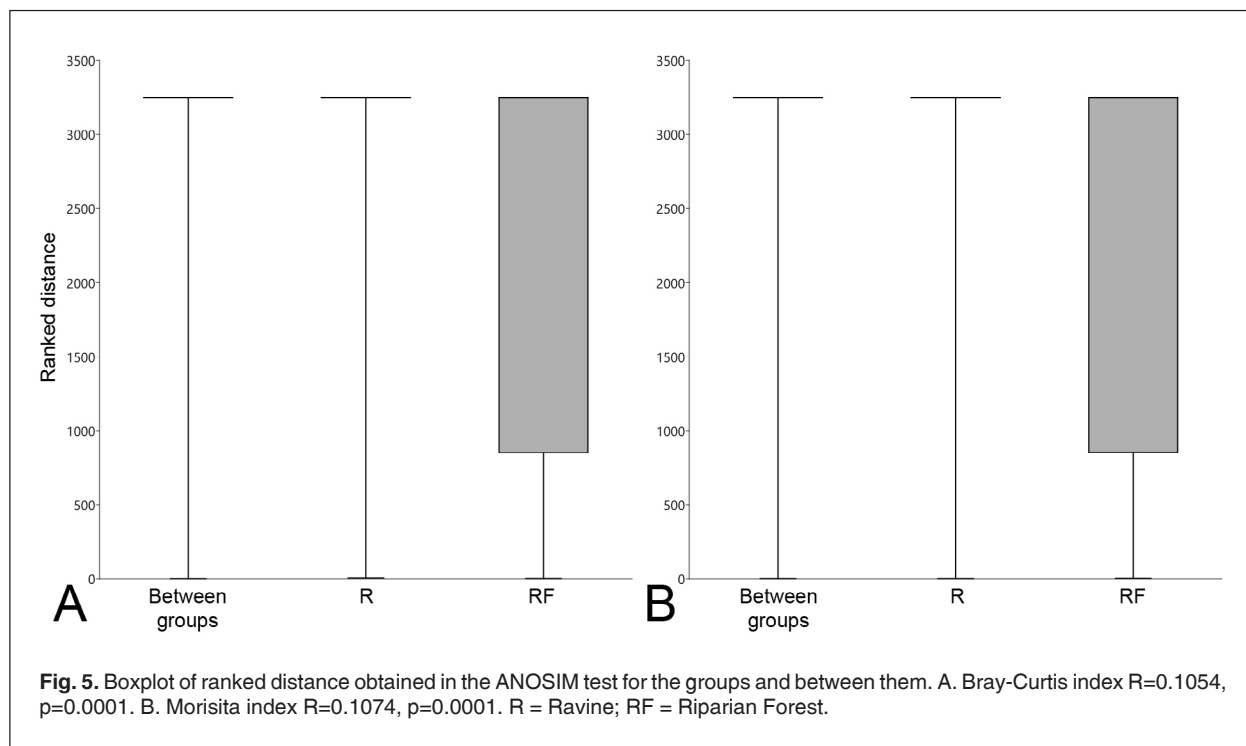


Table 2. Relative abundance of spiders (%) and number of species (S) per guild in the two environments.

	Riparian Forest		Ravine	
	%	S	%	S
Other Hunters	25	51	19	14
Orb Weaver	24	50	34	25
Ground Hunters	14	28	17	13
Space Web	21	43	12	9
Sheet Hunters	6	12	8	6
Sheet Web	2	5	3	2
Sensing Web	1	2	3	2
Specialists	1	2	4	3
Ambush Hunters	6	12	0	0

2016; Mun3var *et al.* 2020). Oliveira *et al.* (2015), applying the analysis of the Geographic Interpolation of Endemism, delimited the areas of endemism of spider species for Brazil. Santos *et al.* (2017), using the Kernel density estimator, identified the areas of greatest richness for the family Araneidae in the Neotropical Region. Both studies reported areas of endemism and highest spider species richness in

southern Brazil bordering eastern Uruguay. Considering the high spider species richness found in Paso Centuri3n and Sierra de R3os, this protected area should probably be influenced by the terrestrial connections with these Brazilian hotspots. This topic should be tested in the future. Vegetation heterogeneity provides more microhabitats for webs building (Rodrigues *et al.*, 2014; Rodrigues *et al.*,

2015). In our study, spider guilds constituted a wide variety of strategies where orb-weavers were the richest. This is consistent with the high availability of shelters for the construction of their webs. In addition, the great abundance and diversity of weaver species explains the presence of several kleptoparasitic spiders (e.g. *Argyrodes*, *Ariamnes* and *Faiditus*), which live and feed on the webs of the larger spiders. In the specific case of *Faiditus plaumanni*, its greater abundance in the riparian forest could be explained by the presence of *Trichonephila clavipes* in the same environment, a large orb weaver indicated as a common host of this kleptoparasitic species (Exline & Levi, 1962).

Statistical analysis revealed that both forest types are different in terms of spider composition and the low contribution of species to this dissimilarity indicates that species are more equitably represented. In addition, species replacement between them revealed high beta diversity. For instance, conservation efforts in the area should take into account the conservation plans of the different forest types in which the high species richness and structure of spider community are located.

This study showed that biodiversity assessments in megadiverse groups such as spiders could provide important taxonomic and ecological data that represent tools for conservation management at local and regional scales in protected areas and their biogeographic connections. Future studies should focus on exploring at a regional scale the role of Paso Centurión and Sierra de Ríos as a link between Atlantic Forest ecoregions.

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